



## COMMENTARY

# Encrypted Morphogens of Skeletogenesis

## BIOLOGICAL ERRORS AND PHARMACOLOGIC POTENTIALS

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**ABSTRACT.** Bone morphogenetic proteins (BMPs) are members of a class of ancient, highly conserved signalling molecules that play major roles in embryonic axis determination, organ development, tissue repair, and regeneration throughout the animal kingdom. The bone morphogenetic proteins are potent developmental morphogens that act in a concentration-dependent manner to specify cell fates in developing and regenerating systems. Complementary DNAs have been cloned for approximately twenty BMPs, and recombinant proteins have been produced for many of these genes. Transgenic and naturally occurring animal models demonstrate a wide variety of potential functions for BMP genes during development and tissue regeneration, and a wide range of pharmacologic effects are predicted from knock-out or over-expression of the BMP genes. Fibrodysplasia ossificans progressiva (FOP), a rare and devastating genetic disease of ectopic osteogenesis in humans, is associated with over-expression of at least one of the BMPs. The BMPs, their transmembrane receptors, their intracellular signal transducers, and their secreted antagonists hold great promise as pharmacologic agents in modulating a vast array of developmental and regenerative pathways in human diseases. *BIOCHEM PHARMACOL* 55:4:373–382, 1998. © 1998 Elsevier Science Inc.

**KEY WORDS.** bone morphogenetic protein (BMP); BMP type I and type II receptors; chordin and noggin; fibrodysplasia ossificans progressiva; heterotopic ossification; morphogen; SMAD proteins

### FIBRODYSPLASIA OSSIFICANS PROGRESSIVA

Rare diseases often provoke important insights into more common disorders and provide a better understanding of normal physiology gone awry. FOP<sup>¶</sup> is an extremely rare, autosomal dominant disorder of connective tissue, with a prevalence of about one case per two million people [1, 2]. Those who have this disorder can be described as essentially forming two skeletons: a normotopic one during embryogenesis and a heterotopic one following birth. The post-natal heterotopic skeleton rigidly immobilizes the joints of the body, rendering movement impossible. Although FOP was first described more than 300 years ago, the genetic defect and pathophysiology remain unknown [2].

FOP is characterized by congenital malformation of the great toes (Fig. 1) and by progressive, disabling heterotopic osteogenesis in distinct anatomic patterns [3–7]. Spontaneous or trauma-induced exacerbations of FOP during childhood are characterized by massive painful soft tissue swelling (Fig. 1) [8]. Histopathologic studies of pre-osseous lesions reveal lymphocytic infiltration and muscle cell

degeneration [9], followed by the appearance of highly vascular, fibroproliferative tissue and finally endochondral ossification with mature lamellar bone and marrow elements [10].

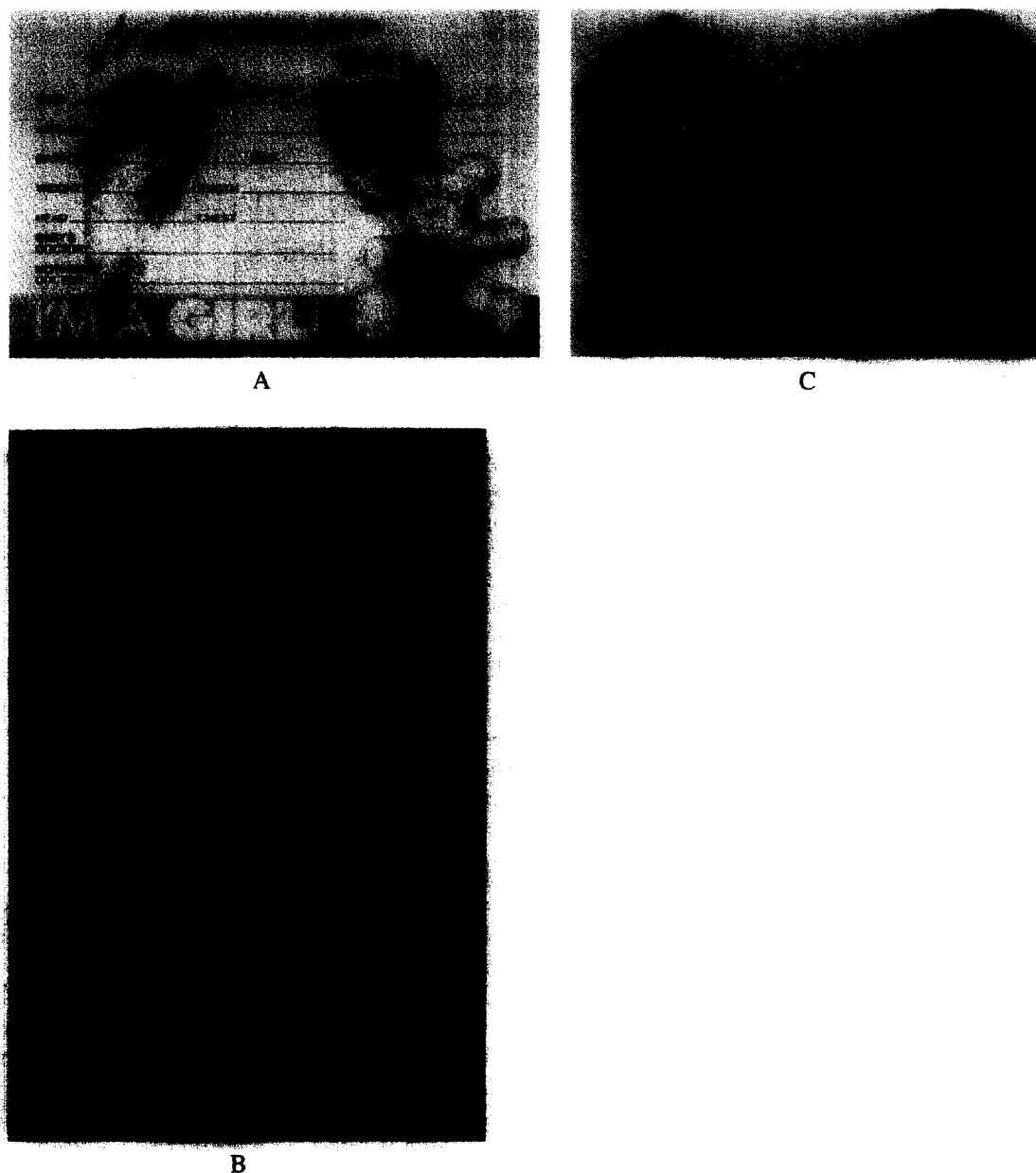
Heterotopic ossification in FOP begins in childhood and can be induced by minor trauma [11]. Severe scoliosis may develop due to asymmetric heterotopic ossification of paravertebral muscles [12]. By early adulthood, heterotopic ossification leads typically to ankylosis of all major joints of the axial and appendicular skeleton, rendering movement impossible [7, 8, 13] (Fig. 2). Most patients are relegated to a wheelchair by their early twenties and require life-long assistance in performing activities of daily living [4, 7]. Starvation and pneumonia are common causes of death [8]. Surgical trauma associated with resection of heterotopic bone leads to exacerbation of local ossification [8]. At present, there is no effective prevention or treatment.

### IDENTIFICATION OF CANDIDATE GENES FOR FOP

The usual approach to identifying the genetic basis of a disease (by genetic linkage analysis and positional cloning) presently is impossible for FOP due to a low reproductive fitness that results in a small number of affected individuals and a lack of multi-generational families showing inheritance of the disease. The candidate gene approach is therefore being pursued as an alternative but indirect method of attempting to identify the mutated gene respon-

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<sup>¶</sup> Abbreviations: BMP, bone morphogenetic protein; FOP, fibrodysplasia ossificans progressiva; TGF transforming growth factor; *dpp*, decapentaplegic; *Osx*, osteoblast-specific transcription factor; *Cbfa*, core binding factor alpha; *CDMP*, cartilage-derived morphogenetic protein; *Gdf*, growth differentiation factor; *AJF*, aggressive juvenile fibromatosis; and mRNA, messenger ribonucleic acid.



**FIG. 1.** Characteristic features of FOP in early childhood. The presence of short malformed great toes at birth (A and C) heralds the later spontaneous appearance of pre-osseous soft-tissue lesions on the neck and back (B) (arrow heads), and should provoke suspicion of FOP even before the transformation to heterotopic bone (arrows). An inspection of the toes will confirm the diagnosis and may alleviate the need for a lesional biopsy (trauma) that could exacerbate the condition. [Reprinted with permission from *J Bone Miner Res* 12: 855, 1997.] Copyright (1997) American Society for Bone & Mineral Research. [Ref. 3].

sible for FOP. In selecting candidate genes for FOP, the main diagnostic criteria (congenital malformations of the great toes with an anterior polarizing defect, heterotopic endochondral ossification, and temporal and spatial patterns of ectopic bone formation) must be considered. The candidate gene for FOP would need to be one that is functional during normal embryonic development (to account for the malformations of the great toe), and one that also could be activated post-natally to induce severe generalized heterotopic ossification in tendon, ligament, fascia, and skeletal muscle.

Genes that plausibly fit these criteria are the *BMP* genes and genes in the *BMP* pathway [14, 15]. *BMP* was the name

initially given to a demineralized bone extract that had the ability to induce ectopic bone formation in an animal assay system [16].

#### **BMPs AND BMP SIGNALLING PATHWAYS**

Morphogens are secreted peptides that diffuse from their source (thus establishing gradients) and act on transmembrane receptors to specify cell fate in a concentration-dependent manner. Substantial data support the role of BMPs as morphogens in animal development [15, 17–23]. Many of the BMPs act as inducers of endochondral osteogenesis and fracture healing [14, 16–21, 24–28]. BMPs are



FIG. 2. Clinical photograph and skeleton of a man with FOP. The rigid posture noted in this 25-year-old man with FOP is due to ankylosis of the spine, shoulders, and elbows. Plates and ribbons of ectopic bone contour the skin over the back and arms (A), and can be visualized directly on the skeleton (B) (following death from pneumonia at age 40 years). Courtesy, Mütter Museum, College of Physicians of Philadelphia. [Reprinted with permission from *N Engl J Med* 335: 555–561, 1996; Fig. 1.] Copyright (1996) Massachusetts Medical Society. All rights reserved. [Ref. 13].

unique in their ability to induce the complete cellular program of endochondral osteogenesis at heterotopic sites *in vivo* [14, 15, 29–31]. Although their family name implies a predominant role in skeletogenesis, the BMP functions include somite development, limb patterning, neural crest lineage determination, and development of the kidneys, teeth, lung, heart, liver, gut, testes, skin, and skeleton [19, 32–38] (Table 1).

Based on similarity of protein structure, the BMPs are part of the larger TGF- $\beta$  superfamily of peptides [15]. BMPs are the most phylogenetically conserved members of the TGF- $\beta$  superfamily and are ubiquitous regulators of animal development [15]. Among the BMPs, BMP-4 plays a vital and seminal role in the induction of all mesoderm [21].

Interestingly, the genes with the highest degree of similarity to members of the mammalian BMP family have been found in the fruit fly, *Drosophila melanogaster* [22, 52]. The BMP-2 and BMP-4 genes, which produce proteins that

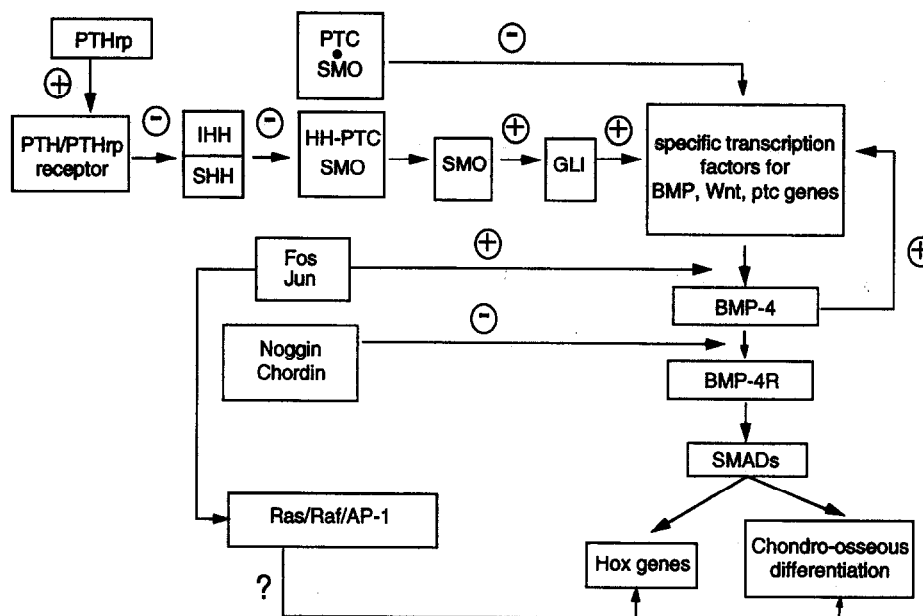
are about 90% similar to each other, are homologous to the *Drosophila dpp* gene. The DPP protein shows ~75% amino acid identity to BMP-2 and BMP-4 in the mature carboxy-terminal region of these proteins [22, 52]. As secreted peptides, they act as dorsalizing gradient morphogens in dipteran development, and as ventralizing gradient morphogens in vertebrate development [15, 53]. The dipteran and vertebrate BMP-4 homologs are able to substitute functionally for each other *in vivo* in genetic complementation studies [22, 54].

Hox genes [18], *hedgehog* genes [55], fibroblast growth factor genes [56], and the Ras/Raf/AP-1 pathway genes are putatively involved in BMP-4 signalling [57]. The developmental action of the BMPs can be modulated *in vivo* by the secreted peptides chordin/sog and noggin [56, 58–65], which have a high binding affinity for the BMPs and competitively inhibit the binding of BMP to its transmembrane receptors [58]. In vertebrate neurogenesis, the BMP

TABLE 1. Established tissue expression for vertebrate bone morphogenetic proteins

BMP	Skf	Ten	Tth	Skn	Msc	Bnm	Hrt	Lng	Liv	Fat	Gut	Kid	Pan	Cns	Eye	Prs	Ovy	Tst	References
BMP-2/dpp	+		+	+			+	+				+					+		[37, 39, 40]
BMP-4/dpp	+		+	+	+		+	+			+	+		+					[17, 39]
BMP-5	+							+	+			+							[39, 41]
BMP-6/Vgr-1	+							+				+							[39, 42, 43]
BMP-7/OP1	+		+	+				+				+			+				[35, 36, 39, 44]
BMP-8a/OP2																		+	[39, 41]
BMP-8b																		+	[32, 39]
CDMP-1/Gdf-5	+																		[26, 39]
BMP-13/Gdf-6	+																		[45]
BMP-12/Gdf-7		+						+				+					+		[46]
BMP-3/osteonin	+							+			+		+						[39, 47]
Gdf-10/BMP-3b	+				+		+	+											[48]
BMP-9									+										[49]
Gdf-1														+					[50]
Gdf-3						+				+									[51]
Gdf-8					+														[51]
Gdf-9																	+		[50]

Expression includes both embryonic and post-natal expression. See Ref. 39 for a general review. Abbreviations: Skf, skeleton (bone or cartilage); Ten, tendon/ligament; Tth, teeth; Skn, skin/hair follicle; Msc, muscle/somites; Bnm, bone marrow/spleen/thymus; Hrt, heart; Lng, lung; Liv, liver; Fat, adipose tissue; Gut, gut/intestine; Kid, kidney; Pan, pancreas; Cns, brain/CNS; Prs, prostate; Ovy, ovary; Tst, testes; OP, osteogenic protein; and Vgr, Vg (vegetal) related.



**FIG. 3.** Putative BMP-4 signalling pathways. This composite schematic diagram depicts experimentally determined features of BMP signalling that have been identified in numerous *in vitro* and *in vivo* model systems including *Drosophila*, *Xenopus*, chicken, mouse, and human. Each component of this pathway may not be active in every model system or in every cell type within a model system. Key: (+) indicates an activating pathway; and (-) indicates an inhibitory pathway. Abbreviations: PTHrP, parathyroid hormone related protein; PTH, parathyroid hormone; IHH, Indian hedgehog; SHH, sonic hedgehog; HH, hedgehog (either IHH or SHH); PTC or ptc, patched; SMO, smoothened; GLI, glioblastoma-derived oncogene family; BMP, bone morphogenetic protein; Wnt, vertebrate wingless family; BMP-4R, receptors for BMP-4; Hox, vertebrate homeobox family; Fos and Jun, members of the AP-1 family of transcription factors.

antagonists noggin and chordin (secreted by the Spemann organizer during early gastrulation in *Xenopus*) inhibit BMP activity, thereby allowing a default neural cell fate to develop rather than an epidermal one [56, 59–65]. Later in neurogenesis, BMPs are expressed selectively in the developing nervous system and play a major role in specifying the fate maps of neural crest cells [66]. Upstream regulators of BMPs have been identified and include the hedgehog family of signalling molecules (sonic hedgehog and Indian hedgehog), which bind to the patched receptor on the cell surface to derepress the activity of the transmembrane protein smoothened. This activity leads to expression of downstream transcription factors and ultimately to induction of BMP expression [67–69] (Fig. 3).

Few downstream targets of BMP activity have been identified. *Osf2/Cbfa1* is a recently cloned and obligate transcriptional activator of osteoblast differentiation regulated by BMP activity. *Osf2/Cbfa1* protein binds to the promoter to regulate the expression of numerous genes expressed in osteoblasts. The spurious expression of *Osf2/Cbfa1* in pluripotent mesenchymal cells and in mouse skin fibroblasts induces the mature osteoblast-specific phenotype. *Osf2/Cbfa1* is positively regulated by BMP-7 and BMP-2 and inhibited by 1,25-dihydroxyvitamin D [70, 71]. Homozygous knockout of the *Osf2/Cbfa1* gene in mice results in complete lack of bone formation in both the endochondral and intramembranous pathways due to a failure of osteoblastic differentiation and a failure of hematopoietic development [71]. Mice heterozygous for the

*Osf2/Cbfa1* deletion exhibit skeletal abnormalities characteristic of the human skeletal disorder cleidocranial dysostosis [72]. Mutations in *Osf2/Cbfa1* cause cleidocranial dysplasia in humans, and heterozygous loss of function is sufficient to produce the phenotype [73]. Conversely, BMP over-expression leads plausibly to over-expression of *Osf2/Cbfa1* and a robust osteogenic phenotype.

BMPs exert their biological effects via heteromeric (type I and type II) serine-threonine kinase transmembrane receptors [74–82]. Despite their ability to bind type I and type II receptors separately, BMPs require the steric interaction of these two receptors for optimal binding and signal transduction [80]. The intracellular effects of the BMPs are transduced by the cytoplasmic SMAD proteins, a class of transcription co-factors that are translocated to the nucleus following BMP-mediated cytoplasmic phosphorylation [77, 83, 84]. Little is known about the immediate downstream molecular targets of BMPs in vertebrate development [85].

Null mutations in the genes of two members of the BMP family have been identified in mammals. Each of these mutations results in highly restricted skeletal abnormalities during embryogenesis. Homozygous deletions of the *BMP-5* gene have been identified in the *short ear* mouse. These mutations lead to malformations of the axial skeleton and also result in abnormal fracture repair [25]. Homozygous mutations in *CDMP-1*, the human homologue of mouse *Gdf-5*, lead to Hunter–Thompson acromesomelic chondrodysplasia, a recessive disorder with skeletal abnormalities restricted to the limbs and synovial joints [86]. Homozygous

mutations of *Gdf-5*, a member of the BMP family, have been identified in the *brachypodism* mouse and result in malformations of the appendicular skeleton [26].

The absence of BMP-4 in a transgenic knockout mouse (genetically engineered to have no functional BMP-4 genes) is lethal in early embryogenesis, showing little or no mesodermal differentiation and no hematopoiesis [20, 21]. BMP-4 has also been implicated in patterning of the developing mouse limb. In addition, over-expression of BMP-4 in the chick embryonic limb bud has been associated with ectopic osteogenesis [87]. Targeted disruption of BMP-4 signalling in the same model leads to defects in apoptosis and failure to form the interdigital spaces [82]. These highly informative BMP mutations provide evidence for a direct role for at least some of the BMPs in embryonic and post-natal bone formation [15]. Thus, BMPs play critical roles in early embryogenesis and in skeletal formation, important criteria in considering them as plausible candidate genes for FOP.

### INVESTIGATING THE CELLULAR AND MOLECULAR BASIS OF FOP

*Drosophila* genetics, mouse genetics, and developmental biology have provided us with several clues toward understanding BMP function and toward selecting the BMPs as reasonable candidate genes for FOP [52]. Studies in our laboratory have examined the expression of many of the BMPs in cells from FOP patients.

Early FOP lesions are histologically indistinguishable from AJF lesions; however, these two disorders can be distinguished by immunohistochemistry with BMP-2/4 antibodies [88]. Cells from AJF lesions (which do not progress to form bone) show no binding of the BMP-2/4 antibody. FOP lesional cells and tissue bind the BMP-2/4 antibody, indicating the presence of BMP proteins within very early FOP lesions [88]. The antibody used for these experiments cannot distinguish between BMP-2 and BMP-4. However, the activity of these two BMP genes can be distinguished by examining specific mRNA expression [13].

Northern analysis and ribonuclease protection assays were performed to specifically examine the expression of BMP-2 and BMP-4 mRNAs in cells from FOP patients [13]. Cells derived from a pre-osseous FOP lesion and from immortalized lymphoblastoid cell lines established from FOP patients show increased expression of BMP-4 but not BMP-2 compared with controls [13]. Correlation of BMP-4 expression with FOP was also observed in a family showing inheritance of FOP: The affected father and three affected children expressed BMP-4, while the unaffected mother did not. Further studies have verified that BMP-4 protein is synthesized in cells from patients who have FOP [13].

We have proposed that over-expression of a BMP gene may be involved in the gain-of-function leading to heterotopic ossification in patients with FOP [52]. Since the half-life of BMP is extremely brief (only a few minutes), it is unlikely that osteogenic-inducing concentrations of

BMP-4 could be achieved at sites of osteogenesis unless the morphogen is delivered to those sites by circulating cells or manufactured at those sites by mesenchymal cells.

We have proposed that lymphocytes capable of expressing BMP-4 circulate in the peripheral blood of patients with FOP, and are recruited to connective tissue sites after soft-tissue injury [89, 90]. Type IV collagen, a primary constituent of the basement membrane of endothelial cells and muscle cells, avidly binds BMP-4 [91], resulting in increased local concentrations. At high concentrations, BMP-4 acts as a morphogen [14–16, 92] capable of up-regulating its own mesenchymal expression [25] and leading to the development of pre-osseous fibroproliferative lesions [14]. The appearance of lymphocytes in the perivascular space of the earliest detectable FOP lesions [9] provides support for the hypothesis that lymphocytes and perivascular cells may be involved in the induction of osteogenesis [93, 94].

Given the evidence of BMP-4 over-expression associated with heterotopic ossification in some patients with FOP, we are currently pursuing several directions of study to understand the exact involvement of BMP-4 in the pathophysiology of FOP. Recent results have indicated that the increased levels of BMP-4 mRNA in FOP cells are due to an increased rate of transcription of the BMP-4 gene [95]. The increased activation of BMP-4 in FOP cells may be due to a mutation within the BMP-4 gene itself or, more likely, to a mutation in another genetic locus that causes over-expression of BMP-4 in the cells of FOP patients. To begin to examine these questions, we are characterizing the structure and function of the human BMP-4 gene in order to understand how the BMP-4 gene is regulated. To determine whether circulating cells can be a plausible delivery vehicle for biologically significant quantities of BMPs, we are making transgenic animals that over-express BMPs in lymphocytes and other mononuclear cells of hematopoietic origin.

### RELATIONSHIP OF BMP-4 SIGNALLING PATHWAYS TO FOP?

At the present time, a direct genetic link of FOP to the BMP-4 gene has not been proven and remains very circumstantial. The genetic mutation(s) in FOP could plausibly reside anywhere in the BMP-4 signalling pathway, or in other signalling pathways that have effects on the level of BMP-4 expression (Fig. 3). Our research, in the broadest sense, involves an analysis of the genetic and molecular pathways that are ectopically activated in patients who have FOP. We believe that this knowledge will also lead to more effective treatments of congenital skeletal disorders and problems in fracture healing.

### PHARMACOLOGIC POTENTIAL OF THE BMPs AND RELATED MOLECULES

The BMP family of morphogens and their developmental antagonists provide a stunning array of recombinant pep-

tides with dramatic potential for pharmacologic use in humans. Studies have already commenced using recombinant BMPs for a wide array of orthopaedic conditions including augmentation of spinal fusions, promotion of osteogenic ingrowth into total joint arthroplasties, bone reconstruction following surgery for skeletal neoplasia, treatment of fracture non-unions, promotion of healing following osteonecrosis, and bone augmentation for dental and craniofacial reconstruction [96–105]. Many studies suggest that specific recombinant BMPs may have potential use in the regeneration of tendons and ligaments, and in treating various disorders of the urogenital and muscular systems [35, 36, 51, 106].

Creative uses of BMP and BMP-pathway technology will likely have important applications in the inhibition of heterotopic ossification in diseases such as FOP and even in more common forms of clinically disabling heterotopic ossification as encountered following closed head trauma, spinal cord injury, or total hip arthroplasty. Soluble BMP receptors, dominant negative receptors, and recombinant BMP antagonists such as chordin and noggin may be promising in binding and physiologically inactivating BMP where it is not needed or wanted [107].

The hope for an effective treatment for FOP has certainly been boosted by the recent discoveries of BMP-4 over-expression in some patients who have the condition. Investigations are underway to identify the cause of this error in over-expression, which may be in the regulatory regions of the BMP-4 gene or in some other gene whose product regulates BMP-4 expression. Already, however, the findings have heightened hopes for an eventual cure for FOP. "With so much being discovered about how the BMPs act," says Brigid Hogan, a developmental geneticist at Vanderbilt University in Nashville, Tennessee, "it might be possible to develop drugs that would block some part of the BMP-4 pathway—and therefore prevent the progression of what is a horrible, nightmare disease" [108].

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## References

1. Connor JM and Evans DA, Genetic aspects of fibrodysplasia ossificans progressiva. *J Med Genet* 19: 35–39, 1982.
2. McKusick VA, Fibrodysplasia ossificans progressiva (FOP). In: *Mendelian Inheritance in Man. Catalogues of Autosomal Dominant, Autosomal Recessive, and X-Linked Phenotypes* (Ed. McKusick VA), Vol. 11. The Johns Hopkins University Press, Baltimore, 1994.
3. Kaplan FS and Smith RM, Clinical vignette: Fibrodysplasia ossificans progressiva (FOP). *J Bone Miner Res* 12: 855, 1997.
4. Cohen RB, Hahn GV, Tabas JA, Peeper J, Levitz CL, Sando A, Sando N, Zasloff M and Kaplan FS, The natural history of heterotopic ossification in patients who have fibrodysplasia ossificans progressiva. A study of forty-four patients. *J Bone Joint Surg Am* 75: 215–219, 1993.
5. Connor JM, Skirton H and Lunt PW, A three generation family with fibrodysplasia ossificans progressiva. *J Med Genet* 30: 687–689, 1993.
6. Kaplan FS, McCluskey W, Hahn G, Tabas JA, Muenke M and Zasloff MA, Genetic transmission of fibrodysplasia ossificans progressiva. Report of a family. *J Bone Joint Surg Am* 75: 1214–1220, 1993.
7. Rocke DM, Zasloff M, Peeper J, Cohen RB and Kaplan FS, Age- and joint-specific risk of initial heterotopic ossification in patients who have fibrodysplasia ossificans progressiva. *Clin Orthop* 301: 243–248, 1994.
8. Connor JM and Evans DA, Fibrodysplasia ossificans progressiva. The clinical features and natural history of 34 patients. *J Bone Joint Surg Br* 64: 76–83, 1982.
9. Kaplan FS, Gannon FH, Shafritz AB, Zasloff MA and Shore EM, Acute lymphocytic infiltration in a very early lesion of fibrodysplasia ossificans progressiva. Abstracts of the Second International Symposium on Fibrodysplasia Ossificans Progressiva. *Calcif Tissue Int* 59: 136, 1996.
10. Kaplan FS, Tabas JA, Gannon FH, Finkel G, Hahn GV and Zasloff MA, The histopathology of fibrodysplasia ossificans progressiva. An endochondral process. *J Bone Joint Surg Am* 75: 220–230, 1993.
11. Lanchoney TF, Cohen RB, Rocke DM, Zasloff MA and Kaplan FS, Permanent heterotopic ossification at the injection site after diphtheria–tetanus–pertussis immunizations in children who have fibrodysplasia ossificans progressiva. *J Pediatr* 126: 762–764, 1995.
12. Shah PB, Zasloff MA, Drummond D and Kaplan FS, Spinal deformity in patients who have fibrodysplasia ossificans progressiva. *J Bone Joint Surg Am* 76: 1442–1450, 1994.
13. Shafritz AB, Shore EM, Gannon FH, Zasloff MA, Taub R, Muenke M and Kaplan FS, Overexpression of an osteogenic morphogen in fibrodysplasia ossificans progressiva. *N Engl J Med* 335: 555–561, 1996.
14. Wozney JM, Rosen V, Celeste AJ, Mitsock LM, Whitters MJ, Kriz RW, Hewick RM and Wang EA, Novel regulators of bone formation: Molecular clones and activities. *Science* 242: 1528–1534, 1988.
15. Kingsley DM, The TGF- $\beta$  superfamily: New members, new receptors, and new genetic tests of function in different organisms. *Genes Dev* 8: 133–146, 1994.
16. Urist MR, Bone formation by autoinduction. *Science* 150: 893–899, 1965.
17. Jones CM, Lyons KM and Hogan BLM, Involvement of bone morphogenetic protein-4 (BMP-4) and Vgr-1 in morphogenesis and neurogenesis in the mouse. *Development* 111: 531–542, 1991.
18. Francis PH, Richardson MK, Brickell PM and Tickle C, Bone morphogenetic proteins and a signalling pathway that controls patterning in the developing chick limb. *Development* 120: 209–218, 1994.
19. Vainio S, Karavanova I, Jowett A and Thesleff I, Identification of BMP-4 as a signal mediating secondary induction between epithelial and mesenchymal tissues during early tooth development. *Cell* 75: 45–58, 1993.
20. Johansson BM and Wiles MV, Evidence for involvement of activin A and bone morphogenetic protein 4 in mammalian mesoderm and hematopoietic development. *Mol Cell Biol* 15: 141–151, 1995.
21. Winnier G, Blessing M, Labosky PA and Hogan BL, Bone

- morphogenetic protein-4 is required for mesoderm formation and patterning in the mouse. *Genes Dev* 9: 2105–2116, 1995.
22. Padgett RW, Wozney JM and Gelbart WM, Human BMP sequences can confer normal dorsal–ventral patterning in the *Drosophila* embryo. *Proc Natl Acad Sci USA* 90: 2905–2909, 1993.
  23. Zecca M, Basler K and Struhl G, Sequential organizing activities of engrailed, hedgehog and decapentaplegic in the *Drosophila* wing. *Development* 121: 2265–2278, 1995.
  24. Kingsley DM, What do BMPs do in mammals? Clues from the mouse short-ear mutation. *Trends Genet* 10: 16–21, 1994.
  25. Kingsley DM, Bland AE, Grubber JM, Marker PC, Russell LB, Copeland NG and Jenkins NA, The mouse *short ear* skeletal morphogenesis locus is associated with defects in a bone morphogenetic member of the TGF $\beta$  superfamily. *Cell* 71: 399–410, 1992.
  26. Storm EE, Huynh TV, Copeland NG, Jenkins NA, Kingsley DM and Lee S-J, Limb alterations in *brachypodism* mice due to mutations in a new member of the TGF $\beta$ -superfamily. *Nature* 368: 639–643, 1994.
  27. Nakase T, Nomura S, Yoshikawa H, Hashimoto J, Hirota S, Kitamura Y, Oikawa S, Ono K and Takaoka K, Transient and localized expression of bone morphogenetic protein 4 messenger RNA during fracture healing. *J Bone Miner Res* 9: 651–659, 1994.
  28. Bostrom MPG, Lane JM, Berberian WS, Missri AAE, Tomin E, Weiland A, Doty SB, Glaser D and Rosen VM, Immunolocalization and expression of bone morphogenetic proteins 2 and 4 in fracture healing. *J Orthop Res* 13: 357–367, 1995.
  29. Shimizu K, Yoshikawa H, Matsui M, Masuhara K and Takaoka K, Periosteal and intratumorous bone formation in athymic nude mice by Chinese hamster ovary tumors expressing murine bone morphogenetic protein-4. *Clin Orthop* 300: 274–280, 1994.
  30. Takaoka K, Yoshikawa H, Hashimoto J, Ono K, Matsui M and Nakazato H, Transfilter bone induction by Chinese hamster ovary (CHO) cells transfected by DNA encoding bone morphogenetic protein-4. *Clin Orthop* 300: 269–273, 1994.
  31. Urist MR, Bone morphogenetic protein: The molecularization of skeletal system development. *J Bone Miner Res* 12: 343–346, 1997.
  32. Zhao GQ, Deng K, Labosky PA, Liaw L and Hogan BL, The gene encoding bone morphogenetic protein 8B is required for the initiation and maintenance of spermatogenesis in the mouse. *Genes Dev* 10: 1657–1669, 1996.
  33. McGrath SA, Esquela AF and Lee SJ, Oocyte-specific expression of growth/differentiation factor-9. *Mol Endocrinol* 9: 131–136, 1995.
  34. Bellusci S, Henderson R, Winnier G, Oikawa T and Hogan BL, Evidence from normal expression and targeted misexpression that bone morphogenetic protein (BMP-4) plays a role in mouse embryonic lung morphogenesis. *Development* 122: 1693–1702, 1996.
  35. Luo G, Hofmann C, Bronckers ALJJ, Sohocki M, Bradley A and Karsenty G, Bmp-7 is an inducer of nephrogenesis, and is also required for eye development and skeletal patterning. *Genes Dev* 9: 2808–2820, 1995.
  36. Dudley AT, Lyons KM and Robertson EJ, A requirement for bone morphogenetic protein-7 during development of the mammalian kidney and eye. *Genes Dev* 9: 2795–2807, 1995.
  37. Lyons KM, Pelton RW and Hogan BL, Organogenesis and pattern formation in the mouse: RNA distribution patterns suggest a role for bone morphogenetic protein-2A (BMP-2A). *Development* 109: 833–844, 1990.
  38. Wilson PA and Hemmati-Brivanlou A, Induction of epidermis and inhibition of neural fate by BMP-4. *Nature* 376: 331–333, 1995.
  39. Hogan BLM, Bone morphogenetic proteins in development. *Curr Opin Genet Dev* 6: 432–438, 1996.
  40. Lyons KM, Pelton RW and Hogan BLM, Patterns of expression of murine Vgr-1 and BMP-2a RNA suggest that transforming growth factor- $\beta$ -like genes coordinately regulate aspects of embryonic development. *Genes Dev* 3: 1657–1668, 1989.
  41. Özkaynak E, Schnegelsberg PNJ, Jin DF, Clifford GM, Warren FD, Drier EA and Oppermann H, Osteogenic protein-2. A new member of the transforming growth factor- $\beta$  superfamily expressed early in embryogenesis. *J Biol Chem* 267: 25220–25227, 1992.
  42. Wall NA, Blessing M, Wright CV and Hogan BL, Biosynthesis and *in vivo* localization of the decapentaplegic-Vg-related protein, DVR-6 (bone morphogenetic protein-6). *J Cell Biol* 120: 493–502, 1993.
  43. Carey DE and Liu X, Expression of bone morphogenetic protein-6 messenger RNA in bovine growth plate chondrocytes of different size. *J Bone Miner Res* 10: 401–405, 1995.
  44. Marker PC, King JA, Copeland NG, Jenkins NA and Kingsley DM, Chromosomal localization, embryonic expression, and imprinting tests for *Bmp7* on distal mouse chromosome 2. *Genomics* 28: 576–580, 1995.
  45. Hattersley G, Hewick R and Rosen V, *In situ* localization and *in vitro* activity of BMP-13. *J Bone Miner Res* 10: S163, 1995.
  46. Wolfman NM, Celeste AJ, Cox K, Hattersley G, Nelson R, Yamaji N, DiBlasia-Smith E, Nova J, Song JJ, Wozney JM and Rosen V, Preliminary characterization of the biological activities of rhBMP-12. *J Bone Miner Res* 10: S148, 1995.
  47. Hino J, Takao M, Takeshita N, Konno Y, Nishizawa T, Matsuo H and Kangawa K, cDNA cloning and genomic structure of human bone morphogenetic protein-3B (BMP-3b). *Biochem Biophys Res Commun* 223: 304–310, 1996.
  48. Song JJ, Celeste AJ, Kong FM, Jirtle RL, Rosen V and Thies RS, Bone morphogenetic protein-9 binds to liver cells and stimulates proliferation. *Endocrinology* 136: 4293–4297, 1995.
  49. Lee SJ, Expression of growth/differentiation factor 1 in the nervous system: Conservation of a bicistronic structure. *Proc Natl Acad Sci USA* 88: 4250–4254, 1991.
  50. McPherron AC and Lee S-J, GDF-3 and GDF-9: Two new members of the transforming growth factor- $\beta$  superfamily containing a novel pattern of cysteines. *J Biol Chem* 268: 3444–3449, 1993.
  51. McPherron AC, Lawler AM and Lee S-J, Regulation of skeletal muscle mass in mice by a new TGF- $\beta$  superfamily member. *Nature* 387: 83–90, 1997.
  52. Kaplan FS, Tabas JA and Zasloff MA, Fibrodysplasia ossificans progressiva: A clue from the fly? *Calcif Tissue Int* 47: 117–125, 1990.
  53. Graff JM, Thies RS, Song JJ, Celeste AJ and Melton DA, Studies with a *Xenopus* BMP receptor suggest that ventral mesoderm-inducing signals override dorsal signals *in vivo*. *Cell* 79: 169–179, 1994.
  54. Sampath TK, Rashka KE, Doctor JS, Tucker RF and Hoffmann FM, *Drosophila* transforming growth factor  $\beta$  superfamily proteins induce endochondral bone formation in mammals. *Proc Natl Acad Sci USA* 90: 6004–6008, 1993.
  55. Bitgood MJ and McMahon AP, *Hedgehog* and *Bmp* genes are coexpressed at many diverse sites of cell–cell interaction in the mouse embryo. *Dev Biol* 172: 126–138, 1995.
  56. Lamb TM and Harland RM, Fibroblast growth factor is a direct neural inducer, which combined with noggin gener-



- ates anterior-posterior neural pattern. *Development* 121: 3627–3636, 1995.
57. Xu R-H, Dong Z, Maeno M, Kim J, Suzuki A, Ueno N, Sredni D, Colburn NH and Kung H-F, Involvement of Ras/Raf/AP-1 in BMP-4 signaling during *Xenopus* embryonic development. *Proc Natl Acad Sci USA* 93: 834–838, 1996.
  58. Sasai Y, Lu B, Steinbeisser H and De Robertis EM, Regulation of neural induction by the Chd and Bmp-4 antagonistic patterning signals in *Xenopus*. *Nature* 376: 333–336, 1995.
  59. Sasai Y, Lu B, Steinbeisser H, Geissert D, Gont LK and De Robertis EM, *Xenopus chordin*: A novel dorsalizing factor activated by organizer-specific homeobox genes. *Cell* 79: 779–790, 1994.
  60. Zimmerman LB, De Jesús-Escobar JM and Harland RM, The Spemann organizer signal noggin binds and inactivates bone morphogenetic protein 4. *Cell* 86: 599–606, 1996.
  61. Re'em-Kalma Y, Lamb T and Frank D, Competition between noggin and bone morphogenetic protein 4 activities may regulate dorsalization during *Xenopus* development. *Proc Natl Acad Sci USA* 92: 12141–12145, 1995.
  62. Valenzuela DM, Economides AN, Rojas E, Lamb TM, Nuñez L, Jones P, Ip NY, Espinosa Rr III, Brannan CI, Gilbert DJ, Copeland NG, Jenkins NA, Le Beau MM, Harland RM and Yancopoulos GD, Identification of mammalian noggin and its expression in the adult nervous system. *J Neurosci* 15: 6077–6084, 1995.
  63. Holley SA, Neul JL, Attisano L, Wrana JL, Sasai Y, O'Connor MB, De Robertis EM and Ferguson EL, The *Xenopus* dorsalizing factor noggin ventralizes *Drosophila* embryos by preventing DPP from activating its receptor. *Cell* 86: 607–617, 1996.
  64. Piccolo S, Sasai Y, Lu B and De Robertis EM, Dorsoventral patterning in *Xenopus*: Inhibition of ventral signals by direct binding of chordin to BMP-4. *Cell* 86: 589–598, 1996.
  65. Moos M Jr, Wang S and Krinks M, Anti-dorsalizing morphogenetic protein is a novel TGF- $\beta$  homolog expressed in the Spemann organizer. *Development* 121: 4293–4301, 1995.
  66. Shah NM, Groves AK and Anderson DJ, Alternative neural crest cell fates are instructively promoted by TGF $\beta$  superfamily members. *Cell* 85: 331–343, 1996.
  67. Marigo V, Davey RA, Zuo Y, Cunningham JM and Tabin CJ, Biochemical evidence that patched is the Hedgehog receptor. *Nature* 384: 176–179, 1996.
  68. Chen Y and Struhl G, Dual roles for patched in sequestering and transducing Hedgehog. *Cell* 87: 553–563, 1996.
  69. Alcedo J, Ayzenzon M, Von Ohlen T, Noll M and Hooper JE, The *Drosophila* *smoothed* gene encodes a seven-pass membrane protein, a putative receptor for the hedgehog signal. *Cell* 86: 221–232, 1996.
  70. Ducy P, Zhang R, Geoffroy V, Ridall AL and Karsenty G, *Osf2/Cbfa1*: A transcriptional activator of osteoblast differentiation. *Cell* 89: 747–754, 1997.
  71. Komori T, Yagi H, Nomura S, Yamaguchi A, Sasaki K, Deguchi K, Shimizu Y, Bronson RT, Gao Y-H, Inada M, Sato M, Okamoto R, Kitamura Y, Yoshiki S and Kishimoto T, Targeted disruption of *Cbfa1* results in a complete lack of bone formation owing to maturational arrest of osteoblasts. *Cell* 89: 755–764, 1997.
  72. Otto F, Thornell AP, Crompton T, Denzel A, Gilmour KC, Rosewell IR, Stamp GWH, Beddington RSP, Mundlos S, Olsen BR, Selby PB and Owen MJ, *Cbfa1*, a candidate gene for cleidocranial dysplasia syndrome, is essential for osteoblast differentiation and bone development. *Cell* 89: 765–771, 1997.
  73. Mundlos S, Otto F, Mundlos C, Mulliken JB, Aylsworth AS, Albright S, Lindhout D, Cole WG, Henn W, Knoll JHM, Owen MJ, Mertelsmann R, Zabel BU and Olsen BR, Mutations involving the transcription factor *CBFA1* cause cleidocranial dysplasia. *Cell* 89: 773–779, 1997.
  74. Dewulf N, Verschueren K, Lonnoy O, Moren A, Grimsby S, Vande Spiegle K, Miyazono K, Huylebroeck D and Ten Dijke P, Distinct spatial and temporal expression patterns of two type I receptors for bone morphogenetic proteins during mouse embryogenesis. *Endocrinology* 136: 2652–2663, 1995.
  75. Ishidou Y, Kitajima I, Obama H, Maruyama I, Murata F, Imamura T, Yamada N, Ten Dijke P, Miyazono K and Sakou T, Enhanced expression of type-I receptors for bone morphogenetic proteins during bone formation. *J Bone Miner Res* 10: 1651–1659, 1995.
  76. Liu F, Ventura F, Doody J and Massagué J, Human type II receptor for bone morphogenetic proteins (BMPs): Extension of the two-kinase receptor model to the BMPs. *Mol Cell Biol* 15: 3479–3486, 1995.
  77. Liu F, Hata A, Baker JC, Doody J, Cárcamo J, Harland RM and Massagué J, A human Mad protein acting as a BMP-regulated transcriptional activator. *Nature* 381: 620–623, 1996.
  78. Letsou A, Arora K, Wrana JL, Simin K, Twombly V, Jamal J, Staehling-Hampton K, Hoffmann FM, Gelbart WM, Massagué J and O'Connor MB, *Drosophila* dpp signaling is mediated by the *punt* gene product: A dual ligand-binding type II receptor of the TGF $\beta$  receptor family. *Cell* 80: 899–908, 1995.
  79. Nohno T, Ishikawa T, Saito T, Hosokawa K, Noji S, Wolsing DH and Rosenbaum JS, Identification of a human type II receptor for bone morphogenetic protein-4 that forms differential heteromeric complexes with bone morphogenetic protein type I receptors. *J Biol Chem* 270: 22522–22526, 1995.
  80. Ruberte E, Marty T, Nellen D, Affolter M and Basler K, An absolute requirement for both the type II and type I receptors, *punt* and *thick veins*, for dpp signaling *in vivo*. *Cell* 80: 889–897, 1995.
  81. Brummel TJ, Twombly V, Marqués G, Wrana JL, Newfeld SJ, Attisano L, Massagué J, O'Connor MB and Gelbart WM, Characterization and relationship of dpp receptors encoded by the *saxophone* and *thick veins* genes in *Drosophila*. *Cell* 78: 251–261, 1994.
  82. Zou H and Niswander L, Requirement for BMP signaling in interdigital apoptosis and scale formation. *Science* 272: 738–741, 1996.
  83. Hoodless PA, Haerry T, Abdollah S, Stapleton M, O'Connor MB, Attisano L and Wrana JL, MADR1, a MAD-related protein that functions in BMP2 signaling pathways. *Cell* 85: 489–500, 1996.
  84. Graff JM, Bansal A and Melton DA, *Xenopus* Mad proteins transduce distinct subsets of signals for the TGF $\beta$  superfamily. *Cell* 85: 479–487, 1996.
  85. Lecuit T, Brook WJ, Ng M, Calleja M, Sun H and Cohen SM, Two distinct mechanisms for long-range patterning by decapentaplegic in the *Drosophila* wing. *Nature* 381: 387–393, 1996.
  86. Thomas JT, Lin K, Nandedkar M, Camargo M, Cervenka J and Luyten FP, A human chondrodysplasia due to a mutation in a TGF $\beta$  superfamily member. *Nature Genet* 12: 315–317, 1996.
  87. Francis-West PH, Richardson MK, Bell E, Chen P, Luyten F, Brickell P, L. W and Archer CW, The effect of overexpression of BMP-4 and GDF-5 on the development of limb skeletal elements. *Trans Orthop Res Soc* 21: 62–11, 1996.
  88. Gannon FH, Kaplan FS, Olmsted E, Finkel GC, Zasloff M and Shore E, Bone morphogenetic protein (BMP) 2/4 in early fibromatous lesions of fibrodysplasia ossificans progressiva. *Hum Pathol* 28: 339–343, 1997.
  89. Springer TA, Traffic signals for lymphocyte recirculation

- and leukocyte emigration: The multistep paradigm. *Cell* **76**: 301–314, 1994.
90. Buring K, On the origin of cells in heterotopic bone formation. *Clin Orthop* **110**: 293–302, 1975.
  91. Reddi AH and Cunningham NS, Initiation and promotion of bone differentiation by bone morphogenetic proteins. *J Bone Miner Res* **8** (Suppl 2): S499–S502, 1993.
  92. Katagiri T, Yamaguchi A, Komaki M, Abe E, Takahashi N, Ikeda T, Rosen V, Wozney JM, Fujisawa-Sehara A and Suda T, Bone morphogenetic protein-2 converts the differentiation pathway of C2C12 myoblasts into the osteoblast lineage. *J Cell Biol* **127**: 1755–1766, 1994.
  93. Einhorn TA, Majeska RJ, Rush EB, Levine PM and Horowitz MC, The expression of cytokine activity by fracture callus. *J Bone Miner Res* **10**: 1272–1281, 1995.
  94. Brighton CT, Lorch DG, Kupcha R, Reilly TM, Jones AR and Woodbury RA, The pericyte as a possible osteoblast progenitor cell. *Clin Orthop* **275**: 287–299, 1992.
  95. Olmsted EA, Liu C, Haddad JG, Shore EM and Kaplan FS, Characterization of mechanisms controlling bone morphogenetic protein-4 message expression in fibrodysplasia ossificans progressiva. *J Bone Miner Res* **11**: P294, 1996.
  96. Bostrom M, Lane JM, Tomin E, Browne M, Berberian W, Turek T, Smith J, Wozney J and Schildhauer T, Use of bone morphogenetic protein-2 in the rabbit ulnar nonunion model. *Clin Orthop* **327**: 272–282, 1996.
  97. Boyne PJ, Animal studies of application of rhBMP-2 in maxillofacial reconstruction. *Bone* **19**: 83S–92S, 1996.
  98. Cook SD, Baffes GC, Wolfe MW, Sampath TK and Rueger DC, Recombinant human bone morphogenetic protein-7 induces healing in a canine long-bone segmental defect model. *Clin Orthop* **301**: 302–312, 1994.
  99. Cook SD, Baffes GC, Wolfe MW, Sampath TK, Rueger DC and Whitecloud TS III, The effect of recombinant human osteogenic protein-1 on healing of large segmental bone defects. *J Bone Joint Surg Am* **76**: 827–838, 1994.
  100. Cook SD, Wolfe MW, Salkeld SL and Rueger DC, Effect of recombinant human osteogenic protein-1 on healing of segmental defects in non-human primates. *J Bone Joint Surg Am* **77**: 734–750, 1995.
  101. Cook SD and Rueger DC, Osteogenic protein-1: Biology and applications. *Clin Orthop* **324**: 29–38, 1996.
  102. Muschler GF, Hyodo A, Manning T, Kambic H and Easley K, Evaluation of human bone morphogenetic protein 2 in a canine spinal fusion model. *Clin Orthop* **308**: 229–240, 1994.
  103. Riley EH, Lane JM, Urist MR, Lyons KM and Lieberman JR, Bone morphogenetic protein-2: Biology and applications. *Clin Orthop* **324**: 39–46, 1996.
  104. Schimandle JH, Boden SD and Hutton WC, Experimental spinal fusion with recombinant human bone morphogenetic protein-2. *Spine* **20**: 1326–1337, 1995.
  105. Yasko AW, Lane JM, Fellingner EJ, Rosen V, Wozney JM and Wang EA, The healing of segmental bone defects, induced by recombinant human bone morphogenetic protein (rh-BMP-2). A radiographic, histological, and biomechanical study in rats. *J Bone Joint Surg Am* **74**: 659–670, 1992.
  106. Inada M, Katagiri T, Akiyama S, Namika M, Komaki M, Yamaguchi A, Kamoi K, Rosen V and Suda T, Bone morphogenetic protein-12 and -13 inhibit terminal differentiation of myoblasts, but do not induce their differentiation into osteoblasts. *Biochem Biophys Res Commun* **222**: 317–322, 1996.
  107. Graff JM, Embryonic patterning: To BMP or not to BMP, that is the question. *Cell* **89**: 171–174, 1997.
  108. Roush W, Protein builds second skeleton. *Science* **273**: 1170, 1996.